

RNA Contamination Removal Magnetic Beads (gDNA Purification)

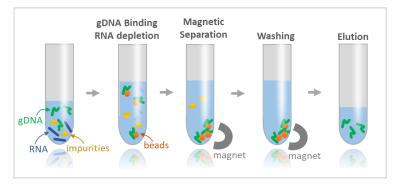
Catalog No.	40060S	40060L
Runs*	40 runs	200 runs

^{*}Based on sample volume of 100 µl

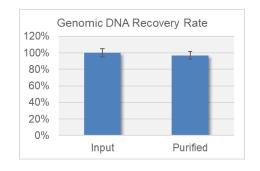
Description

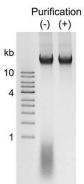
Genomic DNA extraction is the most popular laboratory technique. However, it is common that the extracted genomic DNA is contaminated with RNA. To reduce the impact on downstream applications, it is critical to purify the genomic DNA by removing RNA contamination.

We have developed a simple reagent to completely remove RNA contamination in the extracted genomic DNA samples based on Solid Phase Reversible Immobilization (SPRI) beads. The magnetic beads are coated with carboxyl groups that can reversibly bind to DNA in an appropriate buffer. Our **RNA Contamination Removal Magnetic Beads** combines the reversible DNA-binding properties of SPRI magnetic beads with BioDynami's unique chemistries. The reagent removes the contaminated RNA and recovers the genomic DNA in the same reaction step. Moreover, unwanted components such as proteins, dNTPs, salts, enzymes, and other impurities can also be removed in the same step.



The genomic DNA can be used for downstream applications such as PCR, QPCR, sequencing, microarray, and enzymatic treatment etc. The beads can be an effective and inexpensive reagent for RNA depletion for routine genomic DNA purification.





Features

- Effective removal of RNA contamination by RNase
- High recovery rate of genomic DNA by magnetic beads
- Simple and fast beads-based protocol
- Removal of unwanted components and impurities



Component

Catalog No.	40060S	40060L
RNA Contamination Removal Magnetic Beads	40 runs	200 runs

Storage Condition

Store at 4°C, stable up to 12 months.

Reagent & Equipment Needed (not provided in this reagent)

- Magnetic particle concentrator
- 96-well PCR plate or 1.5 ml tubes
- Thermal cycler, incubator, or heat block
- 80% ethanol (prepare before use)

Protocol

Note: Invert or shake the bottle to thoroughly resuspend the beads.

- 1) Transfer samples to a 96-well plate or 1.5 ml tubes, and add 0.5X volumes of the beads to the wells containing samples. Slow pipetting of the viscous beads is needed for precise aliquot. Mix by pipetting gently and thoroughly.
- Incubate at 37°C for 10 min. Note: Incubation time can be extended in case of heavy RNA contamination.
- 3) Load the sample plate on a magnet, incubate for 3 min, and discard the supernatant carefully.
- 4) Add 180 µl of 80% ethanol without disturbing the beads. Incubate for 2 min and discard the supernatant carefully. Remove all residual ethanol without disturbing the beads. Repeat 80% ethanol wash one more time.
- 5) Air-dry the beads on the magnet for 1-2 min.
- 6) Remove the plate from the magnet and resuspend the beads in at least 20 μl of water, Low TE Buffer or TE Buffer. **Note**: Resuspending the beads in less than 20 μl will reduce the yield.
- 7) Load the plate on the magnet, incubate for 1 min, and transfer the supernatant (containing DNA) to a new tube without disturbing the beads.



Quality Control

Magnetic beads components passed stringent functional quality test.

Product Use Limitation

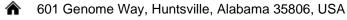
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